Reactions of Thiols and Selenols with the Fe-Mo-S Cluster $[Fe(MoS₄)₂]$ **³⁻**

JOHN W. McDONALD*, G. DELBERT FRIESEN** and WILLIAM E. NEWTON[†] *Battelle-Kettering Laboratory, Yellow Springs, Ohio 45387, U.S.A.* (Received March 24, 1986)

Abstract

The known complex [Et,N] 3 [Fe(Mo&)J has Im known complex $\text{[EqN]}_3\text{[Fe(M034)]}_2\text{]}$ has been shown by EPR and visible spectral studies to react with both thiophenol and selenophenol. The reaction results in a change in the characteristic $S = 3/2$ EPR spectrum of this species from a complex rhombic pattern to one of a very simple axial appearance. Although this effect is similar to that observed for reaction of these species with the ironmolybdenum cofactor of nitrogenase, a moiety known to consist of a $Fe-Mo-S$ cluster species, the large excesses of reagents and the long reaction times required for complete formation of product indicate that these reactions are of questionable direct relevance to the biological system. The reaction corresponding to the EPR spectral change from rhombic to axial in the $[Fe(MoS₄)₂]$ ³⁻/PhSeH system has also been partially characterized by product isolation which indicates that attack by selenol of the two terminal $MoS₂$ moieties in the starting material has occurred.

Introduction

The iron-molybdenum cofactor (FeMoco) is the μ extruded included in the non-morphement conactor (remoco) is the extruded, putative N_2 -reducing site of nitrogenase $[1]$, an Fe-Mo-S cluster obtained by acid treatment of the molybdenum-iron protein of that enzyme, followed by neutralization and extraction with the organic solvent N-methylform amide (NMF) $[2-4]$. The dithionite-reduced form of both the molybdenum-iron protein and FeMoco exhibits an unusual EPR signal at low temperature which is characteristic of $S = 3/2$ magnetic ground state behavior with g values at about 4.5, 3.5 and 2.0 $[5]$. The similarity of the signal of the native protein to that of the ex-

ructural cluster is structure that I choco shares in the entire entire end of FeMoco in $\frac{1}{2}$ in the state state state indicates that $\frac{1}{2}$ is $\frac{1}{2}$ in an analog for the $\frac{m}{L}$ mulcates that symmesis of an analog for the numerous and the begins have been made to simulate the μ and σ are μ and σ and σ spectrum properties α FeMore via synthetic studies of Γ_{α} of Γ_{α} and γ clusters of FeMoco via synthetic studies of Fe-Mo-S cluster
complexes [6]. One such species is $[Fe(MoS₄)₂]$ ³⁻, $\frac{1}{2}$ one such species is $\frac{1}{2}$ (mosq) in no. m_{eff} and m_{eff} model complex, which, and m_{eff} way a structural model for FeMoco, does exhibit an EPR signal characteristic of an $S = 3/2$ ground state $[7-9]$. The EPR signal of FeMoco is known to sharpen markedly on the addition of one equivalent of
thiol [10–11] or selenol [12], becoming more $\frac{100}{100}$ $\frac{10-11}{100}$ or setched $\frac{12}{100}$, becoming more $\frac{d}{dx}$ in general appearance to that of the mory σ denum-iron protein. As part of our overall efforts to study various aspects of Fe-Mo-S clusters for
prototypic behavior for the molybdenum site in nototypic behavior for the morybuenum site in phogenase, we have studied the effect of the phenol and selenophenol on the EPR spectrum of $[Fe(MoS₄)₂]$ ³⁻ and herein report our results.

truded cluster is strong evidence that FeMoco shares

Experimental

Materials and Methods

eriuis una methods
Die carried out in degas de solvents All Icactions were carried out in degassed sorrenes under an atmosphere of pure argon using standard Schlenk tube techniques. $[Et_4N]_3 [Fe(MoS_4)_2]$ was synthesized by the literature method [9] and thiophenol (Aldrich) and selenophenol (Alfa) were used as received. Acetonitrile (MeCN) was distilled from CaH₂. Infrared spectra were obtained on a Beckman IR-20A instrument and visible spectra on a Cary 118C spectrophotometer. EPR spectra were record-
ed using a Varian 4502 spectrometer equipped with a using a varian 4502 spectrometer equipped with $\frac{1}{2}$ and $\frac{1}{2}$ a and an X-band microwave bridge. Samples were
cooled with liquid helium boil-off using either ooled with hour hendin bou-on using children \mathbf{u} and \mathbf{u} dependent $\sum_{i=1}^{n}$ collier voltaments were carried out

with a three-three-electrode cell using MeCN as solvent and with a three-electrode cell using MeCN as solvent and $0.1 \,$ M $[Bu_4N]$ $[BF_4]$ (Aldrich) as supporting electro-

^{*}Author to whom correspondence should be addressed. *Author to whom correspondence should be addressed.

^{**}Present address: Corning Glass Works, Corning, N.Y. 14831, U.S.A. $+9.5$. Western Regional Research Center, \mathbf{R}

Present address: western Regional

lyte. The working electrode was a glassy carbon disk from Biom Bioanalytical Systems (BAS), the reference electronic electronic electronic elecom bioanalytical systems (DAS) , the fereforce electrode was an aqueous SCE separated from the sample solution by a salt bridge, and the potentiostat was a CV-1A unit from BAS. For low temperature work, the cell was immersed in a dewar cooled to -40° C with an acetone-dry ice bath.

Visible Spectral Studies

 10^{-4} to 10^{-3} M solutions of $\left[\text{Et}_4\text{N}\right]_3\left[\text{Fe(MoS}_4)_2\right]$ were placed in either a 4 ml-capacity of a 0.3 mlcre placed in chirch $a + \text{im}$ -expacity of a 0.5 m capacity cell, which had pathlengths of 1.0 cm and 0.1 cm , respectively, and were equipped with septa to et al. Alice and well equipped with septa p_{sc} in \mathbf{M} contains into the reaction into the reaction tenor solutions in meets were injected more reacat anxieties and the resulting spectra monitored recorded. In a time course of the time course of selected. In a time course of selection, the time course of s $\frac{1}{2}$ cording and $\frac{1}{2}$ recording spectra at $\frac{1}{2}$ spectra at $\frac{1}{2}$ reactions was monitored by recording spectra at various intervals after addition of the reagent.

EPR Spectral Studies

Spectral Staties
 $\frac{1}{2}$ reaction mixtures were removed in 3 mm ID α action inixidies were removed, placed in β min TD spectral and those in highly introgen for fact of the reaction was more than the reaction was more than the course visible spectrum was informed and correlated to the visible spectral change. In other cases, the semiquantitative rate of the reaction, determined from separate tive rate of the reaction, determined from separate t_{S} spectral studies, was used to assure that read- $\frac{1}{2}$ complete before the same from the store from the store $\frac{1}{2}$ and $\$ complete before the samples were frozen and stored in liquid nitrogen.

Preparative Scale Reaction of [Et4N],(Fe(MoS4),] withing with PhSeH
The complex $(0.20 \text{ g}; 0.22 \text{ nmol})$ was dissolved in

 M_{\odot} mecomplex (0.20 g, 0.22 millor) was dissolved in $\frac{1}{2}$ and the reaction mixture was added by syringe. After 2 h, the reaction mixture was filtered and the filtrate evaporated under vacuum to $ca. 5$ ml. Addition of $Et₂O(45$ ml) produced a tacky $b \cdot b$ in Addition of Eq. (4) hill product a tacky (28.6) was isolated with the Filtration, which with $\frac{1}{28.6}$ (0.28 g) was isolated by filtration, washed with Et₂O, and dried *in vacuo. Anal.* Found: C, 38.83; H, 4.89; $\frac{1}{2}$ and $\frac{1}{2}$ at the control of the crystallization of the crystallization of the crystallization of the crystal cr $f(z, z, \tau)$. Attempted feerystamization of this product from $MeCN/Et_2O$ gave only powdery samples which
had almost identical IR and electrochemical data and μ annost fuentical IN and electrochemical data and entical El K spectra to those of the initially isolated solids. However, typical elemental analytical data for these 'recrystallized' samples were significantly different (Found: C, 34.68; H, 4.32; N, 2.25%) from that of the crude product and difficulty was encountered in obtaining consistent C , H , and N analyses in general. In addition, neither the overall sulfur and selenium analyses nor the S/Se ratio were consistent from sample to sample (Product A: S, 10.75 ; Se, 20.78 . Product B: S, 7.75 ; Se, 23.39).

Results and Discussion

Since its synthesis several years ago $[7-8]$, the properties of the $\frac{13}{2}$ in $\frac{13}{$ operies of the $[Fe(m)34/2]$ foll (1) have been investigated extensively, largely because a Fe-Mo-S moiety which exhibits an $S = 3/2$ EPR signal, similiar to that found in 1, is known to be present in nitrogenase $[5]$, and, as noted above, is thought to comprise the substrate-reducing site of this important enzyme [1]. Although X-ray absorption spectroscopic (XAS) experiments have conclusively shown $[6]$ that 1 is in no way a realistic structural model for FeMoco (the synthetic cubane-type $Fe₃MoS₄$ clusters $[13-14]$ mimic the XAS properties of the biological unit much more closely), nevertheless the magnetic properties of this complex in particular have been studied as they relate to those of the biological system. EPR [9] and Mössbauer [15] spectroscopy studies of 1 have suggested that the net three unpaired electrons associated with 1 arise from antiferromagnetic coupling between a single Fe(III) and two $Mo(V)$ atoms. Recent magnetic susceptibility and magnetization data $[16]$, however, have been interpreted in terms of the presence of $Fe(I)$ in 1. Thus, this complex has served as a useful prototype for characterization of spin coupling
phenomena in heterometallic sulfur-containing heterometallic sulfur-containing clusters. Because of our general interest in the spectroscopy to 1 and because changes in the EPR spectrum of FeMoco were used to demonstrate [10, 12] and quantitate $[11]$ the reaction of thiols and selenols with this $S = 3/2$ Fe-Mo-S cluster, we have studied the reactivity of these reagents with 1 , using EPR spectroscopy as the primary monitor, but also characterizing the system with other spectral and electrochemical measurements and with limited preparative studies, in order to attempt to define the nature of the products. $\frac{1}{\sqrt{1-\frac{1$

Tigue I shows the ETR spectral changes enched on addition of large excesses of thiophenol (PhSH) and selenophenol (PhSeH) to a solution of $[Et_4N]_3$ -

g. 1. EPR spectra of a ca . 10 \overline{a} M solutions of 1 in MeCN which contain no reagent (top), 1.5 M PhSH (middle), and 1.5 M PhSeH (bottom). $T = 10$ K; microwave frequency = 9.15 GHz.

Fig. 2. Visible spectra of a $ca. 10^{-4}$ M solution of 1 in MeCN containing no reagent $(- - -)$, 1.5 M PhSH $(-)$, and 1.5 M PhSeH (------).

 $[Fe(MoS₄)₂]$ (1) in MeCN. It is evident that the S = 3/2-type spectral pattern characteristic of **1** is greatly simplified in the presence of these reagents, changing from a complex, rhombic-like appearance [9] to one which is almost perfectly axial with g values at 4.01 and 2.03. The concomitant visible spectral changes associated with this reactivity are shown in Fig. 2, with the characteristic peak pattern of 1 replaced by single broad absorption bands in the 450-500 nm region for both PhSH and PhSeH. The similarity of the final EPR and visible spectra for these two reactants suggests that the overall stoichiometry of the products for these two systems is identical except for the S/Se dichotomy.

In order to obtain the complete conversion of 1 to products, an excess of thiol or selenol was required. If less than a ca. 750 fold excess of PhSH or a 20 fold excess of PhSeH was added at the low concentrations of **1** used for spectral studies, both EPR and visible spectra of reaction mixtures at infinite time showed the presence of both reactant and product. As shown in Figs. 3 and 4, the reaction of 1 with PhSH was relatively slow as monitored by both EPR and visible spectroscopy. The visible spectral data was obtained in the presence of a 600-fold

Fig. *3.* Time course of the visible spectral change for reaction of a $ca. 10^{-3}$ M solution of 1 in MeCN with 0.61 M PhSH: (1) 0 min; (2) 60 min; (3) 300 min; (4) 1000 min.

Fig. 4. Time course of the EPR spectral change for reaction of a $ca. 10^{-3}$ M solution of 1 in MeCN with 1.5 M PhSH: (a) 0 min; (b) 3 min; (c) 15 min; (d) 300 min. $T = 10$ K; microwave frequency = 9.15 GHz.

excess of reagent and required ca. 1000 min before absorbance changes ceased. Even then, as noted above, the reaction had not gone to completion. The rate of the analogous reaction of **1** with PhSeH was much faster than for PhSH, being effectively instantaneous at a 600-fold excess of reagent. From visible spectral studies using lower concentrations of PhSeH, we estimate that the rate for the selenol is $ca.$ 10 to 20 times faster than that for the thiol. It is also evident from Fig. 3 that the conversion of 1 to product is chemically 'clean', based on the presence of several isosbestic points in the visible spectral change as a function of time. In keeping with this observation, it should be noted that the overall integrity of the $S = 3/2$ spin system in 1 is clearly preserved in the products, even under the rather harsh conditions involving very high concentrations of PhSH and PhSeH and long reaction times. Thus, reaction other than simple redox behavior or decomposition of **1** is indicated in these systems.

Attempts to isolate the product from the l/PhSH system met with no success, apparently due to the large excess of thiol required for complete reaction. Evaporation of reaction mixtures yielded only oily residues which resisted solidification on treatment with a variety of solvents. Better results were obtained on reaction of **1** with PhSeH where, as noted above, less reagent is required for complete formation of product. Thus, after solvent removal and trituration of the residue with $Et₂O$, a dark brown solid (2) was isolated from a MeCN reaction mixture containing an 18 fold excess of PhSeH. Attempts to purify this product by recrystallization were not particularly successful, yielding only powders whose carbon, hydrogen, nitrogen, sulfur and selenium

analytical data were not consistent from sample to sample and were different from the data for the initially-isolated solid. Thus, although the data positively confirmed the presence of selenium in PhSeHtreated **1,** only limited information about the stoichiometry of 2 could be gleaned from elemental analysis. The presence of varying amounts of extraneous PhSeH in 2 could produce these inconsistent analytical data, but we have no evidence to confirm this possibility. Fortunately, EPR and visible spectra of solutions of this product in MeCN were virtually identical to those described above. So, even though the solid product is somewhat impure, information obtained from characterization of 2 thus is likely to be relevant to the product generated *in situ* from **1** plus excess PhSeH and, because of the spectral similarities, almost certainly to the product from reaction of **1** with PhSH.

Fig. 5. Cyclic voltammograms of *ca.* 10^{-3} M solutions of 1 (bottom) and 2 (top) in MeCN at -40° C. Scan rate = 50 mV/s.

Figure 5 shows reductive-scan cyclic voltammograms of 2 and its precursor **1** as obtained in MeCN at -40 °C. Conversely, electrochemical measurements on 2 at ambient temperatures showed an additional broad wave at $ca. -1.1$ V which increased in intensity with time, as well as decreased reversibility of the redox event at $ca. -1.30$ V, thus indicating decomposition of 2 at the higher temperature under electrochemical conditions. Two features from the voltammograms are worthy of note. First, the trace for 2 is fairly clean, containing only weak oxidation waves between -0.3 and 0.0 V in addition to the two major waves. This indicates either that the complex is relatively pure or that any major impurities, in 2 are not redox active in this voltage range. Second, the basic redox pattern of **1** (a reversible one-electron reduction at -1.70 V and an irreversible one-electron oxidation at -0.07 V) is conserved in 2, but is shifted by $ca. 350$ mV to more posive potentials. The redox events in 1 have prevusly been assigned to $[Fe(M_0S_4)_2]^{2-3}$ and $[Fe (M_0S_4)_2$]^{3-/4-} couples. The similarity of the voltarnmograms would seem to be consistent with the conservation of the basic $Mo-S_2-Fe-S_2-Mo$ framework in 2, although the evidence is obviously equivocal. In addition, it may be noted that the shift of the redox events of this postulated framework to more positive potentials is in keeping with the addition of 'softer' ligands (perhaps PhSe- or PhSeH) to the electrochemically active unit. For example, a similiar positive shift was noted for the $2-\frac{3}{-}$ couple in the heterotrinuclear complexes $[Co(WO_x S_y$, $\frac{1}{2}$ ^{-/3-} as oxygen was replaced by sulfur [17].

 \mathbf{F} , 6. Infrared spectra of 1 (right) and 2 (left) as KBr pellets.

Figure 6 shows the low-energy portion of the infrared spectra of the isolated solids **1** and 2. On reaction with PhSeH. the characteristic strong pattern of terminal and bridging MO-S and MO-S-Fe bands in the $400-500$ cm⁻¹ region is replaced by a single broader band at 475 cm^{-1} . A number of bands due to the phenyl vibrations of PhSeH are also evident in the $650-750$ cm⁻¹ region. The spectrum in the M-S region is reminiscent of that displayed by the $[Cl_2FeS_2MoS_2FeCl_2]$ ²⁻ ion, a linear trinuclear species [18] which contains $[MoS₄]$ ²⁻ chelating two $FeCl₂$ molecules and thus has no terminal MO-S linkages. Certainly the spectral change in Fig. 6 is strong evidence that reaction of **1** with selenophenol (and by inference thiophenol also) in some way involves attack on the two terminal ∞ moieties in the starting material. The presence f the 475 cm^{-1} band, which is reasonably assigned to a Fe $-S_2$ -Mo bridge vibration, is also a good indication (and one that complements the above observations from EPR and electrochemical studies) that some sort of heteronuclear Fe-MO-S framework remains in 2.

Conclusions

Thiophenol and selenophenol react with [Fe(Mo- S_4)₂]³⁻ to yield new Fe-Mo-S clusters which are structurally similar based on their EPR and visible spectral properties. The reactions are relatively slow and require an excess of reagent for complete conversion to product. Characterization of the product from the $[Fe(MoS₄)₂]³$ -/PhSeH system by elemental analysis, infrared spectroscopy, and cyclic voltammetry are consistent with attack by the reagent on the terminal $MoS₂$ moieties of the starting material while retaining the integrity of the basic $S = 3/2$ magnetic unit. Unfortunately, efforts to characterize completely the stoichiometry of this product were unsuccessful. While the overall sharpening of the EPR spectrum of $[Fe(MoS₄)₂]³⁻$ on addition of thiol or selenol is similar in nature to that produced by treatment of the iron-molybdenum cofactor of nitrogenase with these reagents $[10-11]$, the large excesses of reactants and the long reaction times required for formation of products would seem to indicate that these reactions are not directly relevant to those exhibited by the biological system. However, this synthetic system has provided a starting point for studying the reactivity of these reagents with heterometallic clusters which contain both terminal and bridging sulfide atoms, species in which there is a good deal of current interest [191.

Acknowledgement

This work was supported by grants 81-CRCR-l-0675 (to J.W.M. and W.E.N.) and 85-CRCR-1-1639 (to J.W.M.) from the USDA/SEA Competitive Research Grants Office. This manuscript constitutes Contribution No. 882 from the Battelle-Kettering Laboratory.

References

- 1 T. R. Hawkes, P. A. McLean and B. E Smith, *Biochem. J. R. Hawkes, F. P.*
J.J. 315 (1994). *2*, *21*, *21*, *21*, *21*, *11* (1904).
- *74, 3249* (1977).
- *3 S.-S. Yang,* W.-H. Pan, G. D. Friesen, B. K. Burgess, J. L. Corbin, E. I. Stiefel and W. E. Newton, J. *Biol. Chem., 257, 8042 (1982).*
- *4* M. J. Nelson, M. A. Levy and W. H. Orme-Johnson, Proc. *Nat. Acad. Sci. U.S.A., 80, 142* (1983).
- 5 B. K. Burgess, in C. Veeger and W. E. Newton (eds.), A. Duigess, in C. Veeger and W. E. Newton (eds.) W Junk, The Hanguer and Pudoc, Wageningen, 1984, p. \sim 6 R. H. Holm, *Chem. Sot. Rev., 10, 455* (1981); D.
- Coucouvanis, *Act. Chem. Rex, 14, 201* (1981); B. A. Averill, *Strut. Bonding (Berlin), 53, 59* (19833. *Averill, Struct. Bonding (Berlin), 53, 59 (1983).*
7 J. W. McDonald, G. D. Friesen and W. E. Newton, *Inorg.*
- *Chim. Acta, 46,* L79 (1980).
- *8* D. Coucouvanis, E. D. Simhon and N. C. Baenziger, J. *Am. Chem. Sot., 102. 6644* (1980).
- 9 G. D. Friesen, J. W. McDonald, W. E. Newton, W. B. Euler and B. M. Hoffman, *Inorg. Chem., 22, 2202* (1983).
- 10 J. Rawlings, V. K. Shah, J. R. Chisnell, W. J. Brill, R. Zimmerman, E. Munck and W. H. Orme-Johnson, .I *Biol. Chem., 253,* 1001 (1978).
- *Chem., 253, 1001 (1978).*
11 B. K. Burgess, E. I. Stiefel and W. E. Newton, J. Biol. Chem., 255, 353 (1980).
- 12 B. K. Burgess and W. E. Newton, in A. Muller and W. E. Newton (eds.), 'Nitrogen Fixation: The Chemical-Bio- $\frac{1}{2}$ Constitution Interface interface $\frac{1}{2}$ $\ddot{\ }$.
- p. 1.
13 T. E. Wolff, P. P. Power, R. B. Frankel and R. H. Holm, *J. Am. Chem. Sot., 102, 4694* (1980).
- 14 G. Christou, C. D. Garner, F. E. Mabbs and T. J. King, J. *Chem. Sot.. Chem. Commun.. 740* (1978). J , Chem, Boc., Chem, Commun., $140(1370)$.
- $\sum_{n=1}^{\infty} P_n$ $\sum_{n=1}^{\infty} P_n$. $\sum_{n=1}^{\infty} P_n$. Simbon and P. Petrouleas, D. Coucouvanis, E. D. Simhon and P. Stremple, *Chem. Phys. Lett.*, 81, 261 (1981).
- 16 G. A. Bowmaker, P. D. W. Boyd, R. J. Sorrenson, C. A. Reed and J. W. McDonald, *Inorg. Chem.,* submitted for publication.
- 17 A. Muller, R. Jostes, W. Hellman, C. Romer, H. Bogge, U. Multel, R. Jostes, W. Hemilall, C. Romer, H. Dogge, Donald and W. E. Newton, 2. *Anorg. Allg. Chem., 533,* Donald and W. E. Newton, *Z. Anorg. Allg. Chem., 533,* 125 (1986).
- 18 D. Coucouvanis, N. C. Baenziger, E. D. Simhon, P. Stremple and D. Swenson,J. *Am. Chem. Sot., 102, 1732 (1980).*
- 19 A. Muller, E. Diemann, R. Jostes and H. Bogge, *Angew. Chem., Int. Ed. Engl., 20, 934* (1981).